

International Journal of Environment and Climate Change

12(10): 338-347, 2022; Article no.IJECC.87025 ISSN: 2581-8627 (Past name: British Journal of Environment & Climate Change, Past ISSN: 2231–4784)

Status of Genomic Research in Non-basmati Aromatic Joha Rice of Assam

Nayanmoni Borah^a, Ramendra Nath Sarma^b, Vedant Vikrom Borah^a and Partha Pratim Behera^{b*}

^a Assam Don Bosco University, Guwahati, 782402, Assam, India. ^b Department of Plant Breeding and Genetics, Assam Agricultural University, Jorhat, 785013, Assam, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJECC/2022/v12i1030805

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/87025

Mini-review Article

Received 27 February 2022 Accepted 07 May 2022 Published 10 May 2022

ABSTRACT

Many natively adapted, aromatic, and high-quality rice landraces can be found in NE India. Among the different classes of cultivated rice varieties, Assam has a distinctive scented rice variety known locally as '*joha*,' which is particularly popular among the region's farmers. The aroma of *joha* rice is also thought to differ markedly from that of basmati rice. Genotype stability can be established through genetic variation. Thus, new disease resistant *joha* rice varieties are necessary to meet both domestic and worldwide demand. There is a need for conservation of this traditional aromatic joha rice germplasm which could be used in the future breeding programme for multiple disease resistance and grain and aroma quality traits improvement. The recent advancements in marker and genomics assisted breeding coupled with high-throughput genotyping and phenotyping enable the discovery of novel alleles and gene targets for the implementation of sustainable rice cultivars with improved grain aroma quality.

Keywords: Joha rice; aroma quality; marker assisted breeding; genomics assisted breeding.

1. INTRODUCTION

Among cereal crops, rice is a popular choice. Rice is a staple food for more than two-thirds of the world's population [1]. The genus Oryza and the tribe Oryzeae belong to the Gramineae family, which includes rice (Poaceae). The Oryza genus has 25 known species. There are 23 wild

*Corresponding author: E-mail: parthapratimbehera44@gmail.com;

species, two of which are cultivated: *O. sativa* and *O. glaberrima*. It has been suggested that *O. sativa* is the more commonly cultivated of the two cultivated species of cannabis [1,2,3]. It is cultivated all over the world, including Asia, North and South America, the European Union, the Middle East, and Africa. *On* the other hand, *O. glaberrima* is only grown in West Africa.

2. ORIGIN

The origin and diversity of two cultivated species, O. sativa and O. glaberrima, have been documented combining genetic diversity, historical and archaeological evidence, and distribution. The geographical Himalayan foothills, Chhattisgarh, Orissa's Jeypore Tract, north-eastern India, northern Myanmar and Thailand, and Yunnan Province, China are among the Asian cultural hotspots. The Inner Niger Delta and the Guinean coast of Africa are regarded as the African species of O. glaberrima diversity hotspots [4]. The progenitors of O. sativa are O. barthii and O. longistaminata, and those of O. glaberrima are O. barthii and O. longistaminata [4,5]. India is a major rice producer. Indica rice was believed to have been initially domesticated in the foothills of the Eastern Himalayas (north-eastern India), before being introduced to Japan. Assam and Nepal have perennial wild rice [6,7].

3. RICE GENOME

Oryza has 12 basic chromosomes. O. sativa, O. *glaberrima*, and 14 wild species have 24 chromosomes each, whereas eight wild species have 48 chromosomes each. The nine different genomes A, B, C, D, E, F, G, H, and J were established through chromosome pairing behaviour and fertility in interspecific hybrids. Rice is the first cereal crop to have its whole genome sequenced. On average, the genome is 400-430 Mb in size [6].

4. CULTIVATION

Most people in the globe eat rice, which is one of the most important sources of energy. Globally, indica occupies between 85% and 90% of total land area [7]. For example, the Northeast is a rice biodiversity hotspot [8]. Rice is grown in upland, lowland, and deep water. This area has 10,000 indigenous rice cultivars [9]. With 44.6 million hectares of land dedicated to rice cultivation, India leads the world in terms of area. Rice yields around 90 million tonnes and yields 2086 kg/ha. Assam has a large rice cultivar collection. Rice is grown on 25.3 million hectares and produces 3.8 million tonnes and 1540 kg/ha. (Source: Directorate of Agriculture, Govt of Assam, and Guwahati 2009, unpublished data).

5. AROMATIC RICE OF ASSAM

"Specialy Rice" refers to a variety of rice genotypes with specific purposes around the world [10]. Basmati rice, aromatic non-basmati rice, non-aromatic basmati rice, and nonaromatic special purpose rice are all examples of specialty rice [11]. Assam is known for its aromatic, glutinous, and semi-glutenous joha, bora, and chokuwa rice varieties. Joha rice appears to be the most valuable of the bunch. The aromatic rice of Assam is a unique class of rice under Sali rice, traditionally known as 'Joha'. North-eastern (NE) India has a distinct group of locally cultivated non-basmati aromatic rice germplasm and quality rice land races [12]. For example, joha, chakhao, and tai are locally adapted cultivars grown in the states of Assam, Manipur, and Mizoram, respectively. Assam Joha rice is well-known for its distinct flavour, fine grain, easy cooking, and great taste, all of which make it a desirable commodity in society [7]. The smell of Joha rice is a key attribute that contributes to its widespread fame [13]. Aromatic rice is a type of rice that is believed to be of the highest quality and sells for a significantly greater price than non-aromatic rice [14]. So, Joha rice occupies a remarkable position in the local markets compared to other classes of rice. However, because of its scent, it is vulnerable to disease and insect onslaughts. The genetic diversity of joha rice is in jeopardy due to increased yields and the commercial value of longer grain forms [15]. Due to their low production, farmers rarely plant these cultivars for commercial purposes. Nearly 5% of the state's 17 lakh acres of sali rice is joha [16]. These photoperiod reactive, tall conventional joha cultivars yield 1.0 t/ha. Also, joha rice is farmed in marginal lands. They have a similar aroma and quality, but have not been extensively explored [17]. Few trials have been conducted to economically develop high-yielding fragrant rice suitable for this region. Assam Joha rice types were clearly unique from other Assam rice varieties, as evidenced by their distinctive grouping in a DNA diversity analysis based on molecular markers by Bhuyan et al. [18]. This mandates the use of DNA markers to identify these joha rice cultivars [19].

6. JOHA RICE CULTIVAR OF ASSAM

In terms of aroma, longevity, grain size and shape, production ability, and other traits, Assam's aromatic rice, locally known as "joha," differs from other aromatic rices. After transplanting seedlings late in the season, Joha rice is produced in water adjacent fields, hilly lands, or on Sali seed beds. Joha occupies around 5% of the Sali rice land; with an average yield of 1–1.5 t/ha [7]. Talukdar et al. cultivated 54 joha varieties. But after clustering, they got the results for 42 varieties, which came under the aromatic joha category.

Table 1. Some of the collected germplasm maintained at Assam Agricultural University
(Talukdar et al. [17])

SI. No.	Variety Name	SI. No.	Variety Name
4	Arab Joha	25	Tulsi Joha
3	Badshabhog	24	Ronga Joha-2
2	Bengoli Joha	23	Kamini Joha
8	Bhaboli Joha	30	Konbogi Joha
1	Boga Joha	22	Kunkuni Joha-1
9	Boga Joha-1	31	Goalporia Joha-2
6	Boga Maniki Modhuri	27	Kon Joha-3
7	Bogi Joha	28	Kola Joha-2
5	Bokul Joha	26	Monipuri Joha-2
18	Jeera Joha	39	Chufon Joha
20	Keteki Joha	41	Bor Sal Joha
10	Kharika Joha	32	Joha
17	Kolajoha new	38	Goalporia Joha-1
12	Koli Joha-2	33	Koli Joha
19	Kon Joha	40	Cheniguti Joha
14	Kunkuni Joha-2	35	Kon Joha-1
15	Maniki Madhuri Joha	36	Ronga Joha-1
16	Monipuri Joha-1	37	Ronga Joha-1
21	NDR6330	42	Boga Tulsi Joha
13	Siali Joha	34	Joha Bora

7. SPECIFICATION AND DESCRIPTION OF JOHA RICE

Physicochemical properties of the grains, growth patterns, habitat areas, and the shape and cooked grain elongation pattern of this particular aromatic rice class set it apart from Basmati rice. The major grain characteristics of Joha rice genotypes are given below:

Table 2. Physiochemica	I properties of	grains of	joha [1	7]
------------------------	-----------------	-----------	---------	----

5.2 - 10.0 mm
1.9- 3.0 mm
2.3- 4.5 (Short bold to medium
Black, white
7-26 g
1.0- 1.4
65-75%
60-65%
Intermediate
10-20%
20- 24%
0.412- 0.773%
0.121 -0.264%
Soft
0.4 - 4.85 ppm
Strong
19-24%

121- 174 cm
8 - 13
135— 165 days
24 — 33 cm
27 — 41 cm
0.8 — 1.3
30 -44 cm2
108 -303
18— 30%

Table 3. Morphological characters of joha rice

Joha rice is a cross between Indica and Japonica. It belongs to the moderate slim or medium wide class with an L: B of less than 3. A grain's length can grow 1.4 times but ripen when cooked. In many ways, Joha is unlike Basmati rice (the other scented rice of India). Basmati has refined grains (greater than 6.5 mm in length), 1.8 times elongation, a rich scent, and a delicate texture. However, Joha grains are smaller (4.7 mm) and have less elongation (1.1 times). Even though they don't have a distinct aroma and aren't soft after cooking [7]. They deliberated on 37 indigenous Joha cultivars, using "Pusa Basmati" and "Kernel local" as check varieties. Some of the features they looked at were husking recovery, milling recovery, head rice recovery, as well as kernel length and breadth. High diversity in head rice recovery, kernel length, breadth, and L/B ratio, elongation, and alkali value. Each Joha cultivar had a different kernel length and different breath measurements, but the L/B ratio was always 3. The highest elongation percentage (110%) was recorded in Koli Joha, followed by Manipuri Joha (107%) and Ronga Joha (Ronga) [7]. Das et al. compared Joha rice to basmati and kasturi. They discovered joha had higher crude protein content than basmati and kasturi (9.17-11.67%). They found genetic variation and environmental influences in the protein, amylase, amylopectin, and mineral content of the grains. A higher concentration of albumin, globulin, and amylase was detected in indigenous Joha cultivars compared to Basmati and Kasturi. According to their research, Kola Joha, Badshabhog, and Keteki Joha have the strongest aromas. The main reason why fragrant rices are studied is their exquisite aroma, which is attributed to 2acetyl-1-pyrroline. This molecule is more potent than other rice volatile components [20]. Ahmed et al. classified Joha cultivars based on scent strength and pyrroline content [21].

8. GENETICS OF AROMA IN AROMATIC RICE

Among the various grain quality parameters, aroma is the most instantly detectable sensory trait. This is why farmers have prioritized it over other traits for decades. The chewing test has been used to pick up aromas for a long time. Following the introduction of the KOH test, more complex tests were developed using gas chromatography mass spectrometry (GCMS) and other sensitive devices to accurately assess fragrance and identify the volatile components that cause it [22]. Sensory modalities and functional markers correlated with polymorphisms that define the fragrance genes can be used to accomplish accurate phenotyping and genotyping [23]. Aromatic hydrocarbons, aldehydes, phenols, alcohols, ketones, and esters are among the more than 500 volatile aromatic compounds (VACs) found in cooked aromatic rice. 2 acetyl-1-pyrroline is the main VAC contributing to scent and is found in aerial regions of plants and deposits throughout seed maturity [24]. It was determined that a single recessive gene (fgr) was responsible for producing aroma in rice plants, which was connected to the RFLP clone RG28 on chromosome 8 (4.5cM genetic distance) [25,26]. Kovach et al. identified the betaine aldehyde dehydrogenase gene (BADH2) linked with the aromatic phenotype through fine and sequence [27]. Genetically. mapping aroma in rice is investigated to be restricted by а maior gene, osBadh2 (also documented as fgr/badh2/ os2AP /osbadh2, LOC_Os08g0424500), which is found as a homozygous recessive gene at chromosome 8 expressed under mutation of an eight-base-pair (8-bp) deletion "GATTAGGC" in exon 7 codes for betaine aldehyde hydrogenase (osBadh2) results in a premature stop codon, leading to the lack of gene's of activity and a buildup of 2Acetvl-1- Pvrroline (2AP) in an allele at the fragrance locus in aromatic genotypes [7]. [28,29], [30,31,32]. Bradbury et al. also found 3 SNPs in the 7th exon of the Badh2 gene [28]. Using aromatic rice and non-fragrant rice as experimental materials, Chen et al. were able to position the aroma gene (osBadh2) at a distance of 252 KB between 20175367 bp and 20386172 bp on chromosome 8 [33]. Later analysis by Shan et al. revealed that Badh2 is 1509 bp long, with 15 exons and 14 introns [34]. On the other hand, distinct aromatic rice types have 7 bp deletions in exon 2 of the Badh2 gene, as well as an 803 bp loss between exon 4 and exon 5 of the Badh2 gene [32], [35]. Furthermore, the Badh2 gene's 5'UTR region and promoter region insertion/deletion/single nucleotide feature mutation sites [35]. Thus, the Badh2 gene allelic variation affects rice grain fragrance. In addition to the Badh2 gene, two QTLs were discovered on the 3rd and 4th chromosomes that affect rice aromatic traits [34]. The Badh1 aene is homologous to the Badh2 gene and may be associated with QTL on the 4th chromosome.

Several tasty and healthy rice cultivars from South and Southeast Asia lack the 8-bp deletion, yet 2AP was identified in both raw and cooked rice. Fitzgerald et al. claimed that another mutation causes 2AP accumulation and not the fgr 8-bp deletion [31]. Most uniform fgr genotypes had 2AP levels similar to aromatic nfgr genotypes, while certain South Asian genotypes had high levels. Chakraborty et al. studied whether any locus other than osBadh2 controls scent in 84 indica rice landraces [37]. The osBadh2 deletion was detected in practically all landraces. But eleven aromatic genotypes, including wild progenitors, lacked the functional characteristic allele. This revealed a second gene, or allele, controlling fragrant rice. Not all aroma-responsible genes have been identified [38], [27], [39].

Rice aroma eminence is also heavily influenced by the farming process as well as environmental factors such as temperature, soil type, abiotic stress, water, CO2, light, salinity, and shading [40], [41]. According to a study, the basmati aromatic rice variety becomes more aromatic and expresses a super aroma while cultivating in the Punjab of India and Pakistan, but is found to be less aromatic in other regions or countries [6]. The grain articulates a super fine aroma when cultivation is done with humidity of about 70%– 80% and a relatively cool temperature in the afternoon (25 °C–32 °C) and night (20°C–25°C)

at the primal and grain filling stages [42]. The iasmine rice (KDML105, a popular aromatic rice variety) is found to be higher in aroma while growing in various dry land areas in the north and north-eastern parts of Thailand than in other countries [43]. Some studies have concluded that the aromatic rice varieties are typically prone to numerous diseases and pests, abiotic stress, and extremely effective against photoperiodism [44], [45], [46]. The internal mechanism of divergence in aroma development at the molecular level in different environmental conditions has not been studied and elucidated completely. They are used as morphological markers to tell rice landraces apart because changes in the environment have the least effect on the quality of the rice.

9. GENOMIC STUDY IN JOHA RICE FOR AROMA AND YIELD

The study of genetic diversity is an important factor for breeders to explore variability. Rice taxonomists prefer to do rapid classification of different taxonomic groups, while breeders are interested in finding important agronomic variation in breeding programmes [47]. In addition to the two main subspecies, Indica and Japonica, five genetically characterised groupings, indica, aus, aromatic, temperate japonica, and tropical japonica, have been discovered with genetic markers [38], [48], [49], [50], [51]. The genetic structure of O. sativa cultivars around the world has been examined a few times, but few studies have been associated with a specific region [48], [52]. An abundance of genetic differences among the native rice cultivars of northeastern India has been discovered by previous studies based on morphology and other agronomic variables [53], [54], [56]; as well as DNA profiling (isozyme, RAPD, ISSR) [18], [56], [57]. The foothills of the Himalayas, which encompass Uttar Pradesh, Bihar, and Nepal's Tarai area, are the hub of scented rice varieties in India [58]. Aromatic rice germplasm spread from this centre of diversity to other Indian states and neighbouring nations, where it was suited to local conditions. Previous research examined the genomic structure of Indian aromatic rice [59], [60], [61]. However, these studies place less emphasis on NE state fragrant rice accessions. Molecular markers with high polymorphism can be used to map genes/QTLs used in molecular breeding for aroma and yield improvement of Joha rice [17]. It smells stronger than other rice volatiles [20]. AAU investigated the inheritance of

morphological traits. Some studies also claim that the aroma is caused by duplicate factors. Roy et al. classified the 107 numbers of aromatic rice of the NE region into three genetically distinct population clusters: P1, joha rice accessions from Assam, P2, tai rice from Mizoram and Sikkim; P3, chakhao rice germplasm from Manipur; and, aromatic rice accessions from Nagaland. Pair-wise FST was found between three groups and ranged from 0.223 (P1 vs P2) to 0.453 (P2 vs P3). A total of 322 alleles were amplified through 40 simple sequence repeat (SSR) markers with an average of 8.03 alleles per locus, with an average gene diversity of 0.67 revealed to understand the diversification of aroma and quality of rice germplasm for exploiting the genetic diversity for rice improvement in N-E India [12]. Talukdar and co-workers screened 155 simple sequence repeat (SSR) markers for genome wide distribution in rice based on the available map information [15], [62], They found 136 primers were highly polymorphic and spread across 12 chromosomes. They selected seven aromatic primers linked to the BAD2 locus for studying the genetic variability. A total of 143 simple sequence repeat markers were analysed to study genetic variations among the 40 Joha and 14 non-Joha rice genotypes. The polymorphism information content of these markers ranged from 0.17 to 0.88 and found an average of 3.7 alleles. They found three distinct groups among 54 rice accessions through the model-based population assignment and dendrogram analysis, leading to a clear distinction of Joha accessions from the Basmati accession. They detected ten characters through trait association of 29 probable markers with a P value of 0.05. This result in high diversity and a strong population structure in Joha rice. The study of QTLs for yield and grain guality can be further utilised for excellent mapping and validation of specific genes to develop DNA-based molecular markers in rice breeding and better allele mining in Joha rice. By crossing Ranjit and Kola Joha, Talukdar et al. discovered quantitative trait loci (QTLs) that contribute to 12 phenotypic features in rice [17]. They employed 102 SSR markers as well as a few fragrance related markers in their study, creating a 1387.9cM linkage map and discovering 24 QTLs, two of which were for grain aroma on chromosome 5 and chromosome 8, respectively. The investigation also revealed that the QTLs of Aro1-BAD2 were detected in an analogous place to the aroma gene of Basmati rice between the two QTLs. The majority of QTLs in this analysis show a variety of partial to over-

dominance effects. sugaesting that the characteristics are complex. These QTL-linked markers might be exploited in future markerassisted breeding research. Saikia et al. investigated the genetic link between fifteen fragrant Joha rice landraces unique to Assam's Upper Brahmaputra Valley and found a total of 110 polymorphic alleles across all landraces using 34 markers, with an average of 3.25 per locus [63]. For each marker, the Polymorphic Information Content (PIC) varied from 0.24 to 0.83, with an average of 0.5. Another study was done by taking 20 Assam indigenous Joha (aromatic) rice cultivars morphologically. biochemically, and molecularly and finding the aromas of Kalijeera, Kunkuni Joha, Kon Joha-5, Manimuni Joha, and Kon Joha-2 were powerful [64]. Across the 66 polymorphic SSR markers, PCR amplified 174 alleles with a mean value of 2.64, and the average PIC value was 0.326, with values ranging from 0.091 to 0.698. Applying UNJ clusterina dependina on Jaccard's coefficients, the 20 cultivars were classified into three groups, each with eight, ten, and two entries. Chetia et al., (2019) used SNP analysis to investigate the activity of the Badh2 gene in 39 Joha accessions and discovered a link between Badh2 gene fragrances in the majority of the Joha germplasm [65]. Furthermore, the aroma of some aromatic Joha germplasms isn't linked to the Badh2 gene, which could be because of other factors [64], [65].

10. CONCLUSION

Genetic variation can be helpful in establishing a stable and biologically potential genotype. Therefore, new potential disease resistant Joha rice varieties need to be developed to satisfy the demand at both the domestic and global level. Organic farming has now been proven to improve the quality of fragrant rice on a global scale. Along with quality, increasing grain output per unit area is critical to meeting the growing population's food production requirement. The study of the wide genetic diversity of Joha rice varieties can be further utilised for future breeding programmes. The use of association mapping can be helpful for identifying the molecular marker for stacking the QTL responsible for aroma and agronomic traits for varietal development. Novel markers can be used for future validation. Interspecific crosses with high yielding varieties have a chance to give a desirable variety with high yield and aroma. Precise genomic and proteomic tools can be used to analyse the biochemical and qualitative characteristics of joha rice. Proper preservation of Joha rice's germplasm is essential for future breeding programmes. Through the use of bioinformatics tools and sequenced data information, speed breeding can be possible. By focusing more attention on the research area of joha rice, it can be beneficial.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Brar DS. Utilization of wild species of genus Oryza in rice improvement. Mono. on Genus Oryza. 2003:283-309.
- 2. Vaughan DA. The wild relatives of rice: a genetic resources handbook. Int. Rice Res. Inst.; 1994.
- Morishima H. Species relationships and the search for ancestor. Biol. of Rice. 1984:3-10.
- 4. Chang TT. The origin, evolution, cultivation, dissemination, and diversification of Asian and African rices. Euphytica. 1976; 25:425-441.
- Siddiq EA, Vemireddy LR, Nagaraju J. Basmati rices: genetics, breeding and trade. Agri. Res. 2012; 1:25-36.
- Peng B, Zuo YH, Hao YL, Peng J, Kong DY, Peng Y, Nassirou TY, He LL, Sun YF, Liu L, Pang RH. Studies on aroma gene and its application in rice genetics and breeding. Jour. of Plant Stud. 2018; 7.
- Das A, Kesari V, Rangan L. Aromatic Joha rice of Assam-a review. Agri. Reviews. 2010; 31:1.
- Myers N, Mittermeier RA, Mittermeier CG, Da Fonseca GA, Kent J. Biodiversity hotspots for conservation priorities. Nature. 2000; 403:853-858.
- 9. Hore DK. Rice diversity collection, conservation and management in northeastern India. Gene. Res. and Crop Evol. 2005; 52:1129-1140.
- 10. Grist D. Rice 5 ed London; 1983.
- 11. Shenoy VV, Kalagudi GM. Amplified fragment length polymorphism in select specialty rice of India. New directions for a diverse planet. In Proceedings of the 4th International Crop Science Congress, Brisbane, Australia 2004.

- 12. Roy S, Banerjee A, Mawkhlieng B, Misra AK, Pattanayak A, Harish GD, Singh SK, Ngachan SV, Bansal KC. Genetic diversity and population structure in aromatic and quality rice (*Oryza sativa L.*) landraces from North-Eastern India. PloS one. 2015; 10:e0129607.
- 13. Nayak AR, Chaudhury D, Reddy JN. Genetic variability, heritability and genetic advance in scented rice. Ind. Agricul. 2002; 46:2.
- 14. Hien NL, Sarhadi WA, Oikawa Y, HIRATA Y. Genetic diversity of morphological responses and the relationships among Asia aromatic rice (*Oryza sativa L*.) Cultivars. Tropics. 2007; 16:343-355.
- 15. Talukdar PR, Rathi S, Pathak K, Chetia SK, Sarma RN. Population structure and marker-trait association in indigenous aromatic rice. Rice Sci. 2017; 24:145-154.
- Sarma RN, Bahar B. Genetic variation of bora rice (glutinous rice) of Assam as revealed by RAPDs. Plant Gene. Reso. Newsletter. 2005; 144:234.
- 17. Talukdar PR, Rathi S, Pathak K, Chetia S, Baruah AR. QTL analysis in Aromatic Rice of Assam, India. J Rice Res. 2017; 5:2.
- Bhuyan N, Borah BK, Sarma RN. Genetic diversity analysis in traditional lowland rice (*Oryza sativa L.*) of Assam using RAPD and ISSR markers. Curr. Sci. 2007; 10:967-972.
- Sakthivel K, Sundaram RM, Rani NS, Balachandran SM, Neeraja CN. Genetic and molecular basis of fragrance in rice. Biotech. Adv. 2009; 27:468-473.
- 20. Buttery RG. 2-Acetyl-1-pyrroline: an important aroma component of cooked rice. Chem. & Ind. 1982; 23:958-959.
- Ahmed SA, Borua I, Sarkar CR, Thakur AC. Volatile component (2-acetyl-1pyrroline) in scented rice. In Proceedings of the Seminar on Problems and Prospects of Agriculture Research and Development in North East India, Assam Agricultural University, Jorhat, India 1995; 55-57.
- 22. Sarkar S, Pattanaik SS, Chattopadhyay K, Chakraborti M, Sanghamitra P, Basak N, Anandan A, Samantaray S, Subudhi HN, Meher J, Kar MK. Genetic Improvement of Rice for Aroma, Nutrition and Grain Quality. ICAR-NRRI.
- 23. Akwero A, Ocan D, Akech W, Lamo J, Ochwo-Ssemakula M, Rubaihayo P. Allelic variations in aroma gene in cultivated rice

varieties. Afri. Crop Sci. Jour. 2020; 28:241-254.

- 24. Ramtekey V, Cherukuri S, Modha KG, Kumar A, Kethineni UB, Pal G, Singh AN, Kumar S. Extraction, characterization, quantification, and application of volatile aromatic compounds from Asian rice cultivars. Rev. in Anal. Chem. 2021; 40:272-292.
- 25. Ahn SN, Bollich CN, Tanksley SD. RFLP tagging of a gene for aroma in rice. Theo. and Appl. Gen. 1992;84:825-828.
- 26. Lorieux M, Petrov M, Huang N, Guiderdoni E, Ghesquière A. Aroma in rice: genetic analysis of a quantitative trait. Theo. and Appl. Gene. 1996; 93:1145-1151.
- Kovach MJ, Calingacion MN, Fitzgerald MA, McCouch SR. The origin and evolution of fragrance in rice (*Oryza sativa* L.). Proceed. of the Nat. Acad. of Sci. 2009; 106:14444-14449.
- Bradbury LM, Fitzgerald TL, Henry RJ, Jin Q, Waters DL. The gene for fragrance in rice. Plant Biot. Jour. 2005; 3:363-370.
- 29. Vanavichit A, Yoshihashi T, Wanchana S, Saengsraku Areekit S. D, W, Kamolsukyunyong Lanceras J. Toojinda T, Tragoonrung S. Positional cloning of Os2AP, the aromatic gene controlling the biosynthetic switch of 2acetyl-1-pyrroline and gamma aminobutyric acid (GABA) in rice. In5th International rice genetics symposium. Manila, Philippines 2006.
- Bourgis F, Guyot R, Gherbi H, Tailliez E, Amabile I, Salse J, Lorieux M, Delseny M, Ghesquière A. Characterization of the major fragance gene from an aromatic japonica rice and analysis of its diversity in Asian cultivated rice. Theo. and Appl. Gene. 2008; 117:353-368.
- Fitzgerald MA, Sackville Hamilton NR, Calingacion MN, Verhoeven HA, Butardo VM. Is there a second fragrance gene in rice? Plant Biotech. Jour. 2008; 6:416-423.
- 32. Shi W, Yang Y, Chen S, Xu M. Discovery of a new fragrance allele and the development of functional markers for the breeding of fragrant rice varieties. Mol. Breed. 2008; 22:185-192.
- Chen S, Yang Y, Shi W, Ji Q, He F, Zhang Z, Cheng Z, Liu X, Xu M. Badh2, encoding betaine aldehyde dehydrogenase, inhibits

the biosynthesis of 2-acetyl-1-pyrroline, a major component in rice fragrance. The Plant Cell. 2008; 20:1850-1861.

- Shan Q, Zhang Y, Chen K, Zhang K, Gao C. Creation of fragrant rice by targeted knockout of the Os BADH 2 gene using TALEN technology. Plant Biotech. Journ. 2015; 13:791-800.
- 35. Ootsuka K, Takahashi I, Tanaka K, Itani T, Tabuchi H, Yoshihashi T, Tonouchi A, Ishikawa R. Genetic polymorphisms in Japanese fragrant landraces and novel fragrant allele domesticated in Northern Japan. Breed. Sci. 2014; 64:115-124.
- Amarawathi Y, Singh R, Singh AK, Singh VP, Mohapatra T, Sharma TR, Singh NK. Mapping of quantitative trait loci for basmati quality traits in rice (*Oryza sativa* L.). Mol. Breed. 2008; 21:49-65.
- Chakraborty D, Deb D, Ray A. An analysis of variation of the aroma gene in rice (*Oryza sativa* L. subsp. indica Kato) landraces. Gene. Res. and Crop Evol. 2016; 63:953-959.
- 38. Huang X, Zhao Y, Li C, Wang A, Zhao Q, Li W, Guo Y, Deng L, Zhu C, Fan D, Lu Y. Genome-wide association study of flowering time and grain yield traits in a worldwide collection of rice germplasm. Nature Gene. 2012; 44:32-39.
- Kaikavoosi K, Kad TD, Zanan RL, Nadaf AB. 2-Acetyl-1-pyrroline augmentation in scented indica rice (*Oryza sativa* L.) varieties through Δ1-pyrroline-5carboxylate synthetase (P5CS) gene transformation. Appl. Biochem. and Biotech. 2015; 177:1466-1479.
- 40. Itani T, Tamaki M, Hayata Y, Fushimi T, Hashizume K. Variation of 2-acetyl-1pyrroline concentration in aromatic rice grains collected in the same region in Japan and factors affecting its concentration. Plant Prod. Sci. 2004; 7:178-183.
- 41. Mo Z, Li W, Pan S, Fitzgerald TL, Xiao F, Tang Y, Wang Y, Duan M, Tian H, Tang X. Shading during the grain filling period increases 2-acetyl-1-pyrroline content in fragrant rice. Rice. 2015; 8:1-10.
- 42. Singh RK, Singh US, Khush GS, Rohilla R. Genetics and biotechnology of quality traits in aromatic rices. Aroma. Rices. 2000:47-70.

- 43. Yoshihashi T, Nguyen TT, Kabaki N. Area dependency of 2-acetyl-1-pyrroline content in an aromatic rice variety, Khao Dawk Mali 105. Jap. Agri. Res. Quarterly: JARQ. 2004; 38:105-109.
- 44. Nadaf AB, Wakte KV, Zanan RL. 2-Acetyl-1-pyrroline biosynthesis: from fragrance to a rare metabolic disease. Jour. Plant Sci. Res. 2014; 1:102.
- 45. Mahajan G, Matloob A, Singh R, Singh VP, Chauhan BS. Basmati rice in the Indian subcontinent: Strategies to boost production and quality traits. Adv. in Agro. 2018; 151:159-213.
- Feng H, Jiang H, Wang M, Tang X, Duan M, Pan S, Tian H, Wang S, Mo Z. Morphophysiological responses of different scented rice varieties to high temperature at seedling stage. Chin. Jour. of Rice Sci. 2019; 33:68-74.
- 47. Zhou Ζ. Characterization of DNA minisatellite sequences from Oryza sativa and their application in DNA L. finaerprintina. Universitv of Missouri-Columbia; 1995.
- Garris AJ, Tai TH, Coburn J, Kresovich S, McCouch S. Genetic structure and diversity in *Oryza sativa* L. Genetics. 2005; 169:1631-1638.
- Caicedo AL, Williamson SH, Hernandez RD, Boyko A, Fledel-Alon A, York TL, Polato NR, Olsen KM, Nielsen R, McCouch SR, Bustamante CD. Genomewide patterns of nucleotide polymorphism in domesticated rice. PLoS Gene. 2007; 3:e163.
- 50. Zhao K, Wright M, Kimball J, Eizenga G, McClung A, Kovach M, Tyagi W, Ali ML, Tung CW, Reynolds A, Bustamante CD. Genomic diversity and introgression in O. sativa reveal the impact of domestication and breeding on the rice genome. PloS one. 2010; 5:e10780.
- Wang CH, Zheng XM, Xu Q, Yuan XP, Huang L, Zhou HF, Wei XH, Ge S. Genetic diversity and classification of *Oryza sativa* with emphasis on Chinese rice germplasm. Heredity. 2014; 11:489-496.
- 52. Glaszmann JC. Isozymes and classification of Asian rice varieties. Theo. and Appl. Gene. 1987; 74:21-30.
- 53. Borkakati RP, Borah P, Deka PC. Genetic divergence in photoperiod-insensitive autumn rice germplasm of northeast

India. In Adv. in Rice Gene. 2003:74-76.

- 54. Vairavan S, Siddiq EA, Arunachalam V, Swaminathan MS. A study on the nature of genetic divergence in rice from Assam and North East Himalayas. Theoretical and Applied Genetics. 1973; 43:213-221.
- 55. Sarma BK, Pattanayak A. Rice Diversity of North East India. Millen. Graph; 2009.
- 56. Glaszmann JC, Benyayer P, Arnaud M. Genetic divergence among rices from Northeast India. Rice Genetics Newsletter.1989; 63.
- 57. Sarma RN, Bahar B. Genetic variation of bora rice (glutinous rice) of Assam as revealed by RAPDs. Plant Genet. Reso. Newsletter. 2005; 144:134.
- 58. Khush GS. Taxonomy and origin of rice. Aro. Rices. 2000:5-13.
- 59. Aggarwal R, Shenoy V, Ramadevi J, Rajkumar R, Singh L. Molecular characterization of some Indian Basmati and other elite rice genotypes using fluorescent-AFLP. Theo. and Appl. Gen. 2002; 105:680-690.
- Jain S, Jain RK, McCOUCH SR. Genetic analysis of Indian aromatic and quality rice (*Oryza sativa* L.) germplasm using panels of fluorescently-labeled microsatellite markers. Theo. and Appl. Gene. 2004; 109:965-977.
- Nagaraju J, Kathirvel M, Kumar RR, Siddiq EA, Hasnain SE. Genetic analysis of traditional and evolved Basmati and non-Basmati rice varieties by using fluorescence-based ISSR-PCR and SSR markers. Proceed. of the Nat. Acad. of Sci. 2002; 99:5836-5841.
- 62. Temnykh S, DeClerck G, Lukashova A, Lipovich L, Cartinhour S, McCouch S. Computational and experimental analysis of microsatellites in rice (*Oryza sativa* L.): frequency, length variation, transposon associations, and genetic marker potential. Geno. Res. 2001; 11:1441-1452.
- Saikia P, Neog B, Gogoi N, Baruah D. Assessment of the Genetic Diversity of Joha Rice Germplasms by using Simple Sequence Repeat Markers. Ind. Jour. of Agri. Res. 2021; 1:7.
- 64. Bordoloi D, Sarma D, Barua NS, Das BK. Morphological, Biochemical and Molecular Characterization of Indigenous Aromatic Rice of Assam.

65. Chetia SK, Hazarika G, Bordoloi S, Talukdar M, Tamuly A, Das BC, Ali S, Ahmed T. Screening of traditional scented

rice (Joha) of Assam for presence of Badh2, blast and brown spot diseases. ORYZA-An Inter. Jour. of Rice. 2019; 56:2.

© 2022 Borah et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/87025